Synthesis of (25R)-5 α -Cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol, a Cytostatic Starfish Steroid[†]

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Received February 13, 1998

The synthesis of (25R)-5 α -cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol (1a), a marine cytostatic steroid, has been achieved in 13 steps (7.8% overall yield) starting from commercially available diosgenin (2). A key step in the synthesis was the dimethyldioxirane oxidation of the enolsilane 16 to introduce the 15α -hydroxy group in the D ring.

Introduction

Polyhydroxysteroids are naturally occurring steroids present in a wide variety of marine organisms.¹ They have been isolated from soft corals, gorgonians, nudibranchs, sponges, and ophiuroids, but starfishes appear to be the richest source of new polyhydroxysteroids.

Although polyhydroxysteroids do not contain stuctural features commonly associated with cytotoxicity, such as alkylation sites, Michael's acceptors, intercalators, or redox-active quinones, some of them show interesting antiproliferative activity in several tumor cell lines.²

Few hypotheses have been made for the activity of steroidal cytotoxins:^{3,4} they may interfere with the eucariotic cell membrane,⁵ they may be enzyme inhibitors,⁶ they may disrupt the cascade of signal transduction.⁷ Despite the interesting bioactivities showed by polyhydroxysteroids, few syntheses of these compounds have been reported.8

Recently^{2a} an investigation of an Antarctic starfish belonging to the Echinasteridae family has led to the isolation of several polyhydroxysteroids and steroidal

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oligoglycosides, some of them displaying cytotoxic or cytostatic activity. In particular three of them (**1a**-**c**), having a 5 α -cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol framework, showed a cytostatic effect upon human bronchopulmunary non-small-cell lung carcinoma cells (NSCLC-N6), by blocking the cell cycle in phase G_1 (period prior the DNA synthesis).



The fact that insufficient amounts of these compounds were available for further pharmacological studies, coupled with the need to evaluate the mechanism of action, prompted us to undertake the synthesis of one of them: (25R)-5 α -cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol (1a).

We envisaged diosgenin (2) as an useful starting material both for its commercial availability and for the presence of functional groups in position suitable for conversion to 1a.

Results and Discussion

The synthesis began with the protection of diosgenin (2) to give the benzyl ether 3 in 90% yield (eq 1). Treatment of compound **3** with $BH_3 \cdot SMe_2$, followed by oxidation with alkaline hydrogen peroxide,⁹ afforded a mixture of two diasteromeric alcohols (4 and 5) in good overall yield (82%) and with a predominance of the transfused 6α -alcohol **4** (OH- 6α (**4**)/OH- 6β (**5**), 10.7/1).

The two diastereomers were separated by flash chromatography on silica gel, and the unambiguous assignment of the stereochemistry of the A/B ring junction was determined by comparison of their CH₃-19¹H and ¹³C NMR resonances (4, trans-A/B ring junction, ¹H NMR δ 0.79 ppm, ¹³C NMR δ 13.3 ppm; **5**, cis-A/B ring junction, ¹H NMR δ 1.15 ppm, ¹³C NMR δ 26.0 ppm).¹⁰

^{*} Authors to whom correspondence should be addressed. Tel.: +39-89-965230. Fax: +39-89-965296. E-mail: guiso@vaxsa.csied.unisa.it. [†] Dedicated to the memory of Professor Luigi Minale, deceased May 11, 1997.

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(25R)-5 α -Cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol



Inversion of the configuration at C-6 of 4 with a twostep procedure (oxidation¹¹ to ketone **6** and subsequent highly diasteroselective reduction with LiAlH₄, eq 2)^{8a} furnished alcohol 7 in 95% overall yield. Compound 7 was first protected as benzyl ether (8, 75% yield) and then subjected to Clemmensen reduction following Seo's procedure¹² to afford (25R)-3 β ,6 β -bis(benzyloxy)-5 α -cholestane-16*β*,26-diol (9, 52%).



Chemoselective silvlation of diol 9 with tert-butyldimethylsilyl chloride (TBDMSCl) and 1,8-diazabicyclo-[5.4.0]undec-7-ene (DBU)¹³ produced the monoprotected derivative 10 (eq 3). The secondary alcohol at C-16 was then easily oxidated with pyridinium dicromate (PDC) to afford ketone 11 in 78% yield (two steps).



Since the target molecule has a 1,2-trans-diol in the D ring, we considered that this moiety could be introduced by opening of a 15,16-epoxide (eq 4). Thus, kinetic enolization of 11 with lithium bis(trimethylsilyl)amide $(LiN(TMS)_2)$ and quenching of the enolate with Nphenyltrifluoromethanesulfonimide¹⁴ (PhN(SO₂CF₃)₂) gave the stable enoltriflate 12. Deoxygenation of the latter with tributyltin hydride (Bu₃SnH) and tetrakis(triphenylphosphine)palladium(0)¹⁵ afforded (25*R*)-26-[(*tert*-butyldimethylsilyl)oxy]- 3β , 6β -bis(benzyloxy)- 5α -cholest-15ene (13) in 47% yield (two steps). Desilylation at C-26 with tetra-n-butylammonium fluoride (TBAF) and subsequent epoxidation with *m*-chloroperbenzoic acid (MCP-BA) furnished compound 15 (31%, two steps). The



oxirane stereochemistry was indicated by the CH₃-18 1,3syn deshielding effect compared to that reported for cholesterol (δ_{H-18} cholesterol, 0.68 ppm; δ_{H-18} **15**, 0.92 ppm)¹⁶ and supported by a ROESY¹⁷ experiment which showed dipolar coupling between the signals at $\delta_{\rm H}$ 3.06 (H-15) and $\delta_{\rm H}$ 3.20 (H-16) with the resonance at $\delta_{\rm H}$ 1.05 (H-14 α). Unfortunately, attempts to open the oxirane ring in acidic conditions resulted in the formation of an uncharacterized mixture of compounds.

We then decided to hydroxylate the ketone 11. Attempts to obtain the α -hydroxy ketone by treatment of the kinetic enolate of 11 with oxodiperoxymolybdenum-

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(pyridine)(hexamethylphosphoric triamide) (MoOPH)¹⁸ were also unsuccessful since the unreacted ketone was always recovered. However highly stereoselective hydroxylation at C-15 was accomplished by a three-step sequence involving the preparation of a hydrolitically labile silyloxy epoxide (eq 5). The applied method¹⁹ started from ketone **11** which was selectively enolized under kinetic control (-78 °C) using the bulky base lithium bis(trimethylsilyl)amide. The regioisomeric enolate was then quenched with chorotrimethylsilane (TM-SCI), giving the silylenol ether **16**. Dimethyldioxirane²⁰ epoxidation and opening of the silyloxy epoxide **17** with camphorsulfonic acid (CSA), gave the C-26 desilylated α -hydroxy ketone **18** in good overall yield (71%).



Assignment of the configuration at C-15 was made on the basis of a ROESY¹⁷ experiment which showed a strong cross peak between the β -axial CH₃-18 (δ 0.88 ppm) and the C-15 proton (δ 3.59 ppm), establishing an α stereochemistry for the C-15 hydroxy group.

The penultimate step of the synthesis was the reduction of the C-16 oxo group (eq 6). This seemingly straightforward reaction was unexpectedly difficult to perform with acceptable stereoselectivity. Several hydride based reagents were used in this step. NaBH₄ and DIBAL-H gave exclusively the undesired cis isomer **19** in 90% and 86% yields, respectively, while NaBH₃CN and LiAlH₄ afforded both **19** and **20** in 1.6/1 (75%) and 1/1.25 (90%) ratio. The BH₃·SMe₂ reduction procedure, according to Brown and Vogel,²¹ afforded exclusively **20** in 58% yield.

Hydrogenolysis (Pd/C) of triol **20** resulted in removal of the benzyl protecting groups to provide (25R)- 5α cholestane- 3β , 6β , 15α , 16β ,26-pentol (**1a**) in quantitative yield. The target compound was identified by comparison of the NMR data with those reported for the natural product.^{2a}

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Work is in progress to evaluate the cytostatic activity of **1a** in vivo.

Experimental Section

General Procedures. Analytical instrumentation and spectral formats are the same as previously described.²² All reactions were carried out under a dry argon atmosphere using freshly distilled solvents unless otherwise noted. Tetrahydrofuran was distilled from sodium and benzophenone. Toluene, methylene chloride, and diethyl ether were distilled from calcium hydride. Glassware was flame dried (0.05 Torr) prior to use. When necessary, compounds were dried in vacuo over P₂O₅ or by azeotropic removal of water with toluene under reduced pressure. Starting materials and reagents purchased from commercial suppliers were generally used without purification. Dimethyldioxirane was prepared according ref 20. Reactions were monitored by thin-layer chromatography (TLC) on Merck silica gel plates (0.25 mm) and visualized using UV light, spraying with H_2SO_4 -Ce $(SO_4)_2$ solution and drying. Reaction temperatures were measured externally. Flash chromatography was performed on Merck silica gel (60, particle size 0.040-0.063 mm). Yields refer to chromatographically and spectroscopically (¹H and ¹³C NMR) pure materials. High-resolution mass spectra (electron impact, EIMS; fast ion bombardment, FIBMS) were obtained at 70 eV and 4 kV (Cs⁺ ion), respectively, on a Fisons VG Prospec mass spectrometer. The NMR spectra were recorded on a Bruker AM-250, Bruker DRX-400, and Bruker DRX-600 spectrometers. Assignments of ¹H and ¹³C NMR resonances were based on DEPT, COSY, HETCOR, and ROESY17 experiments.

(25*R*)-3β-(Benzyloxy)spirost-5-ene (3). To a suspension of NaH (0.43 g, 18 mmol) in THF (5 mL) at 0 °C was slowly added a solution of diosgenin (2, 5.00 g, 12.0 mmol) in THF (35 mL). After the solution was stirred for 0.5 h at 0 °C, benzyl bromide (BnBr, 2.6 mL, 22.0 mmol) and tetrabutylammonium iodide (TBAI, 0.31 g, 0.84 mmol) were added. The resulting mixture was refluxed for 16 h and then quenched with a saturated solution of NH₄Cl (10 mL), concentrated in vacuo to remove the excess THF, and extracted with diethyl ether. The organic phase was dried (Na₂SO₄) and concentrated in vacuo. Crystallization from CHCl₃/MeOH gave **3** (5.45 g, 90%) as white crystals.

3: mp 13[°]4–135 °C; $[\alpha]_D$ –90.5 (c = 1.0, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.79 (3 H, s, Me-18), 0.79 (3 H, d, J = 6.7 Hz, Me-27), 0.98 (3 H, d, J = 6.0 Hz, Me-21), 1.04 (3 H, s, Me-19), 3.28 (1 H, m, H-3), 3.37 (1 H, dd, J = 10.6, 10.6 Hz, H-26), 3.48 (1 H, dd, J = 10.6, 2.9 Hz, H'-26), 4.43 (1 H, bdd, J = 14.6, 6.9 Hz, H-16), 4.56 (2 H, s, CH₂Ph), 5.35 (1 H, m, H-6), 7.31 (5 H, m, C₆H₅); ¹³C NMR (62.5 MHz, CDCl₃) δ 14.5, 16.2, 17.1, 19.4, 20.8, 28.4, 28.7, 30.2, 31.4 (× 2), 31.8, 32.0, 37.0, 37.1, 39.1, 39.7, 40.2, 41.5, 50.0, 56.5, 62.0, 66.8, 69.9, 78.4, 80.7, 109.2, 121.0, 127.3, 127.5 (× 2), 128.3 (× 2), 139.0, 140.9; HR EIMS m/z 504.3633 (calcd 504.3603 for C₃₄H₄₈O₃).

(25*R*)-3β-(Benzyloxy)-5α-spirostan-6α-ol (4) and (25*R*)-3β-(Benzyloxy)-5β-spirostan-6β-ol (5). To a solution of 3 (0.70 g, 1.4 mmol) in THF (10 mL) at 0 °C was slowly added BH₃·SMe₂ (2.8 mL, 2.0 M in THF, 5.6 mmol). After 0.1 h the solution was warmed to room temperature and stirred for a further 20 h. The solution was then cooled at 0 °C, and

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absolute ethanol (2.5 mL), a solution of NaOH (3.5 mL, 3.0 M), and H_2O_2 (0.7 mL, 30% in water) were added in succession. The mixture was refluxed for 1 h, concentrated in vacuo to remove the excess of THF, and extracted with ethyl acetate. The organic phase was dried (Na₂SO₄), filtered, and concentrated in vacuo. The residue was flash chromatographed (silica gel, 5–40% diethyl ether in petroleum ether) to give **4** (0.54 g, 75%) as a white solid and **5** (0.05 g, 7%) as a colorless oil.

4: mp 228–231 °C; $[\alpha]_D$ –48.8 (c = 1.3, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.74 (3 H, s, Me-18), 0.77 (3 H, d, J = 6.7 Hz, Me-27), 0.79 (3 H, s, Me-19), 0.94 (3 H, d, J = 6.0 Hz, Me-21), 3.30 (1 H, m, H-3), 3.34 (1 H, m, H-6), 3.35 (1 H, dd, J = 10.6, 10.6 Hz, H-26), 3.45 (1 H, dd, J = 10.6, 2.9 Hz, H'-26), 4.38 (1 H, bdd, J = 14.6, 6.9 Hz, H-16), 4.50 (1 H, d, J = 11.8 Hz, CHPh), 4.58 (1 H, d, J = 11.8 Hz, CHPh), 7.30 (5 H, m, C₆H₅); ¹³C NMR (62.5 MHz, CDCl₃) δ 13.3, 14.4, 16.3, 17.0, 20.8, 27.9, 28.6 (× 2), 30.1, 31.2, 31.6, 33.7, 36.4, 37.1, 39.6, 40.4, 41.4, 41.6, 51.3, 53.6, 55.8, 61.9, 66.6, 68.9, 69.7, 77.8, 80.5, 109.1, 127.2, 127.4 (× 2), 128.2 (× 2), 138.8; HR EIMS m/z 522.3683 (calcd 522.3709 for C₃₄H₅₀O₄).

5: $[\alpha]_D - 27.9$ (c = 2.5, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.78 (3 H, s, Me-18), 0.78 (3 H, d, J = 6.7 Hz, Me-27), 0.96 (3 H, d, J = 6.0 Hz, Me-21), 1.15 (3 H, s, Me-19), 3.35 (1 H, dd, J = 10.6, 10.6 Hz, H-26), 3.45 (1 H, dd, J = 10.6, 2.9 Hz, H'-26), 3.64 (1 H, m, H-3), 3.69 (1 H, bs, H-6), 4.40 (1 H, bdd, J = 14.6, 6.9 Hz, H-16), 4.45 (1 H, d, J = 11.8 Hz, *CHP*h), 4.55 (1 H, d, J = 11.8 Hz, CDCl₃) δ 14.5, 16.5, 17.1, 20.6, 24.1, 26.0, 28.7, 30.2, 30.6, 31.3, 31.7, 34.5, 34.8, 40.0, 40.2, 40.4, 40.7, 41.6, 44.2, 56.3, 62.2, 66.8, 68.9, 69.5, 72.8, 73.2, 77.2, 80.8, 109.2, 127.3 (× 2), 128.3 (× 2), 139.2; HR EIMS *m*/*z* 522.3745 (calcd 522.3709 for C₃₄H₅₀O₄).

(25*R*)-3β-(Benzyloxy)-5α-spirostan-6-one (6). To a solution of 4 (0.18 g, 0.34 mmol) in CH₂Cl₂ (2 mL) were added 4 Å molecular sieves (m.s., 0.34 g) and PDC (0.26 g, 0.69 mmol). After 2 h the reaction mixture was diluted with diethyl ether (10 mL). Filtration through a short pad of Celite and CaSO₄ (10% in weight) afforded a solution which was concentrated in vacuo to give **6** as a white solid (0.17 g, 95%).

6: mp 178-180 °C; $[\alpha]_D -82.6$ (c = 1.0, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.77 (6 H, s, Me-18, Me-19), 0.78 (3H, d, J = 6.7 Hz, Me-27), 0.97 (3 H, d, J = 6.0 Hz, Me-21), 3.29 (1 H, m, H-3), 3.35 (1 H, dd, J = 10.6, 10.6 Hz, H-26), 3.45 (1 H, dd, J = 10.6, 2.9 Hz, H'-26), 4.40 (1 H, bdd, J = 14.6, 6.9 Hz, H-16), 4.50 (1 H, d, J = 11.8 Hz, CHPh), 4.58 (1 H, d, J = 11.8 Hz, CHPh), 7.32 (5 H, m, C₆H₃); ¹³C NMR (62.5 MHz, CDCl₃) δ 13.1, 14.4, 16.4, 17.1, 21.3, 26.3, 27.9, 28.7, 30.2, 31.5, 36.6, 37.2, 39.5, 40.9, 41.2, 41.5, 46.7, 53.8, 56.4, 56.7, 61.9, 66.8, 69.7, 76.9, 80.4, 109.3, 127.4, 127.5 (× 2), 128.3 (× 2), 138.7, 210. 7; HR EIMS *m*/*z* 520.3544 (calcd 520.3553 for C₃₄H₄₈O₄).

(25*R*)-3β-(Benzyloxy)-5α-spirostan-6β-ol (7). To a solution of **6** (0.15 g, 0.29 mmol) in diethyl ether (15 mL) was added LiAlH₄ (0.43 mL, 1.0 M in THF, 0.43 mmol). The reaction mixture was stirred for 0.2 h and quenched with ethyl acetate (1 mL) and NH₄OH (0.5 mL, 30% aqueous solution). Filtration through a short pad of Celite and concentration in vacuo gave **7** (0.15 g, 100%) as a white solid.

7: mp 194–196 °C; $[\alpha]_D$ –62.3 (c = 1.2, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.77 (3 H, d, J = 6.7 Hz, Me-27), 0.78 (3 H, s, Me-18), 0.95 (3 H, d, J = 6.0 Hz, Me-21), 1.03 (3 H, s, Me-19), 3.35 (1 H, m, H-3), 3.35 (1 H, dd, J = 10.6, 10.6 Hz, H-26), 3.45 (1 H, dd, J = 10.6, 2.9 Hz, H'-26), 3.75 (1 H, bs, H-6) 4.38 (1 H, bdd, J = 14.6, 6.9 Hz, H-16), 4.52 (1 H, d, J =11.8 Hz, CHPh), 4.57 (1 H, d, J = 11.8 Hz, CHPh), 7.32 (5 H, m, C₆H₅); ¹³C NMR (62.5 MHz) δ 14.4, 15.6, 16.4, 17.0, 20.7, 28.1, 28.6, 29.8, 30.1, 31.2, 31.6, 31.9, 35.6, 38.3, 39.6, 39.9, 40.4, 41.4, 47.2, 54.1, 55.9, 62.0, 66.7, 69.7, 71.6, 78.1, 80.6, 109.2, 127.3, 127.4 (× 2), 128.2 (× 2), 138.8; HR EIMS m/z522.3734 (calcd 522.3709 for C₃₄H₅₀O₄).

(25*R*)-3β,6β-Bis(benzyloxy)-5α-spirostane (8). To a suspension of NaH (0.20 g, 8.52 mmol) in THF (1 mL) at 0 °C was added a solution of 7 (1.75 g, 3.35 mmol) in THF (9 mL). After the solution was stirred for 0.5 h, BnBr (1.2 mL, 10.2 mmol) and TBAI (0.09 g, 0.24 mmol) were added. The

resulting mixture was heated at reflux for 16 h, quenched with a saturated solution of NH₄Cl (2 mL), concentrated in vacuo to remove the excess of THF, and extracted with diethyl ether. The organic phase was dried (Na₂SO₄), filtered, and concentrated in vacuo. The residue was flash chromatographed (silica gel, 5-15% diethyl ether in petroleum ether) to give **8** as a white solid (1.55 g, 75%).

8: mp 54–56 °C; $[\alpha]_D$ –69.5 (*c* = 1.8, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.81 (3 H, s, Me-18), 0.82 (3H, d, *J* = 6.7 Hz, Me-27), 1.00 (3 H, d, *J* = 6.0 Hz, Me-21), 1.10 (3 H, s, Me-19), 3.35 (1 H, dd, *J* = 10.6, 10.6 Hz, H-26), 3.39 (1 H, m, H-3), 3.45 (1 H, dd, *J* = 10.6, 2.9 Hz, H'-26), 3.46 (1 H, m, H-6), 4.34 (1 H, d, *J* = 12.2 Hz, C*H*Ph), 4.43 (1 H, bdd, *J* = 14.6, 6.9 Hz, H-16), 4.58 (2 H, bs, C*H*₂Ph), 4.60 (1 H, d, *J* = 12.2 Hz, C*H*Ph), 7.34 (10 H, m, C₆H₃); ¹³C NMR (62.5 MHz, CDCl₃) δ 14.5, 15.7, 16.4, 17.1, 20.8, 28.2, 28.7, 30.2, 30.4, 31.3, 31.7, 32.4, 35.1, 36.0, 38.3, 39.9, 40.6, 41.6, 47.7, 54.4, 55.9, 62.2, (68.8, 69.7, 71.1, 78.4, 79.1, 80.7, 109.1, 126.9 (× 3), 127.3, 127.5 (× 2), 128.1 (× 2), 128.2 (× 2), 139.1, 139.6; HR EIMS *m*/*z* 612.4137 (calcd 612.4179 for C₄₁H₅₆O₄).

(25*R*)-3 β ,6 β -Bis(benzyloxy)-5 α -cholestane-16 β ,26-diol (9). To a suspension of zinc amalgam, freshly prepared from HgCl₂ (0.30 g, 1.10 mmol), and zinc powder (3.20 g, 48.9 mmol) were added **8** (0.094 g, 0.186 mmol), dissolved in absolute ethanol (12 mL), and concentrated hydrochloric acid (3 mL, 37%, aqueous solution). The mixture was heated at reflux for 1.6 h, cooled to room temperature, filtered, and concentrated under reduced pressure. The crude material was diluted with chloroform, washed with water, dried (Na₂SO₄), and evaporated. Flash chromatography of the residue on silica gel (0– 1% MeOH in CHCl₃) gave **9** (0.049 g, 52%) as a white solid.

9: mp 94–96 °C; $[a]_D – 12.1$ (c = 1.6, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.87 (3 H, s, Me-18), 0.90 (3 H, d, J = 6.7 Hz, Me-27), 0.97 (3 H, d, J = 6.0 Hz, Me-21), 1.06 (3 H, s, Me-19), 3.36 (1 H, m, H-3), 3.44 (3 H, m, H-6, H₂-26), 4.32 (1 H, m, H-16), 4.33 (1 H, d, J = 12.2 Hz, CHPh), 4.56 (2 H, s, CH_2Ph), 4.59 (1 H, d, J = 12.2 Hz, -CHPh), 7.34 (10 H, m, C₆H₅); ¹³C NMR (62.5 MHz, CDCl₃) δ 13.3, 15.7, 16.6, 18.1, 20.6, 23.7, 28.2, 29.6, 30.4, 32.4, 33.3, 34.9, 35.6, 35.9, 36.0, 36.6, 38.3, 40.0, 42.5, 47.8, 53.8, 54.5, 61.6, 68.4, 69.7, 71.1, 72.4, 78.4, 79.2, 126.9 (× 3), 127.3, 127.5 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.7; HR EIMS m/z 616.4461 (calcd 616.4492 for C₄₁H₆O₄).

(25*R*)-26-[(*tert*-Butyldimethylsilyl)oxy]-3 β ,6 β -bis(benzyloxy)-5 α -cholestan-16 β -ol (10). To a solution of 9 (0.640 g, 1.04 mmol) in CH₂Cl₂ (5 mL) were added 1,8-diazabicyclo-[5.4.0]undec-7-ene (DBU, 0.28 mL, 1.9 mmol) and TBDMSCl (0.250 g, 1.66 mmol). The reaction was stirred for 1.5 h before being diluted with CH₂Cl₂ (15 mL), sequentially washed with 0.1 M HCl (10 mL) and with a saturated solution of NaHCO₃ (10 mL) and water, dried (Na₂SO₄), and concentrated in vacuo. The residue (0.760 g), a colorless oil, was used in the next step without purification.

10: $[\alpha]_{\rm p} - 11.8 (c = 1.5, CHCl_3); {}^{1}\text{H} NMR (250 MHz, CDCl_3)$ $<math>\delta$ 0.04 (6 H, s, $(CH_3)_2$ -Si), 0.86 (3 H, s, Me-18), 0.89 (9 H, s, $(CH_3)_3$ -C), 0.90 (3 H, d, J = 6.7 Hz, Me-27), 0.97 (3 H, d, J = 6.0 Hz, Me-21), 1.06 (3 H, s, Me-19), 3.40-350 (4 H, m, H-3, H-6 and H₂-26), 4.32 (1 H, m, H-16), 4.32 (1 H, d, J = 12.2 Hz, CHPh), 4.55 (2 H, s, CH_2Ph), 4.57 (1 H, d, J = 12.2 Hz, CHPh), 4.55 (2 H, s, CH_2Ph), 4.57 (1 H, d, J = 12.2 Hz, CHPh), 7.32 (10 H, m, C_6H_5); 13 C NMR (62.5 MHz, CDCl₃) δ -5.3 (× 2) 13.3, 15.7, 16.7, 18.1, 18.4, 20.6, 23.7, 25.6, 26.0 (× 3), 28.2, 29.8, 30.4, 32.4, 33.6, 34.9, 35.8, 36.0, 36.2, 36.5, 38.4, 40.0, 42.5, 47.8, 54.5, 61.5, 68.5, 69.7, 71.1, 72.5, 78.4, 79.2, 126.9 (× 3), 127.3, 127.6 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.8; EIMS m/z 730 (M⁺).

(25*R*)-26-[(*tert*-Butyldimethylsilyl)oxy]-3β,6β-bis(benzyloxy)-5α-cholestan-16-one (11). To a solution of 10 (0.760 g, 1.04 mmol) in CH₂Cl₂ (5 mL) were added 4 Å molecular sieves (1.40 g) and PDC (0.782 g, 2.18 mmol). After 2 h the reaction mixture was diluted with diethyl ether (10 mL). Filtration through a short pad of Celite and CaSO₄ (10% in weight) afforded a solution which was concentrated in vacuo and purified by flash chromatography (silica gel, 5–10% diethyl ether in petroleum ether) to give 11 as a colorless oil (0.59 g, 78% two steps). **11**: $[\alpha]_D$ -55.5 (*c* = 2.6, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.04 (6 H, s, (*CH*₃)₂-Si), 0.81 (3 H, s, Me-18), 0.87 (3H, d, *J* = 6.7 Hz, Me-27), 0.89 (9 H, s, (*CH*₃)₃-C), 0.96 (3 H, d, *J* = 6.0 Hz, Me-21), 1.09 (3 H, s, Me-19), 3.34 (1 H, dd, *J* = 10.9, 6.5 Hz, H-26), 3.36 (1 H, m, H-3), 3.45 (1 H, dd, *J* = 10.9, 5.8 Hz, H'-26), 3.46 (1 H, bs, H-6), 4.37 (1 H, d, *J* = 12.2 Hz, *CHP*h), 4.54 (1 H, d, *J* = 12.2, *CHP*h), 4.56 (2 H, s, *CH*₂Ph), 7.32 (10 H, m, C₆H₅); ¹³C NMR (62.5 MHz, CDCl₃) δ -5.4 (× 2) 13.8, 15.7, 16.7, 18.6, 20.4, 24.6, 25.9 (× 3), 28.2, 29.7, 31.2, 32.3, 33.3, 35.2, 35.7, 35.9, 36.0, 38.1, 38.9, 39.1, 43.3, 47.7, 50.4, 54.3, 68.2, 68.5, 69.8, 71.4, 78.3, 79.2, 126.9 (× 3), 127.1, 127.3, 127.5 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.6, 218.7; EIMS *m*/*z* 728 (M⁺).

(25*R*)-26-[(*tert*-Butyldimethylsilyl)oxy]-3 β ,6 β -bis(benzyloxy)-5 α -cholest-15-en-16-yl Triflate (12). To a solution of **11** (0.250 g, 0.340 mmol) in THF (4 mL) at -78 °C was added LiN(TMS)₂ (0.77 mL, 1.0 M in THF, 0.77 mmol). After 1 h *N*-phenyltrifluoromethanesulfonimide (0.235 g, 0.660 mmol) was added, and after an additional 0.3 h, the reaction mixture was warmed to room temperature. The reaction was then quenched by addition of water, concentrated in vacuo to remove the excess of THF, and extracted with diethyl ether. The organic layer was washed with a saturated solution of NH₄Cl, dried (Na₂SO₄), and concentrated in vacuo. The residue was flash chromatographed (silica gel, 0–10% diethyl ether in petroleum ether) to give **12** as a colorless oil (0.274 g, 94%).

12: $[\alpha]_D - 8.22 \ (c = 2.0, CHCl_3);$ ¹H NMR (250 MHz, CDCl₃) δ 0.08 (6 H, s, $(CH_3)_2$ -Si), 0.91 (3 H, d, J = 6.7 Hz, Me-27), 0.94 (9 H, s, $(CH_3)_3$ -C), 0.96 (3 H, s, Me-18), 1.04 (3 H, d, J = 6.0 Hz, Me-21), 1.12 (3 H, s, Me-19), 3.39 (1 H, m, H-3), 3.40 (1 H, dd, J = 9.7, 6.5 Hz, H-26), 3.44 (1 H, J = 9.7, 5.9 Hz, H'-26), 3.51 (1 H, bs, H-6), 4.41 (1 H, d, J = 12.2 Hz, *CHP*h), 4.59 (2 H, s, *CH*₂Ph), 4.63 (1 H, d, J = 12.2 Hz, *CHP*h), 5.72 (1 H, bs, H-15), 7.33 (10 H, m, C₆H₃); ¹³C NMR (62.5 MHz, CDCl₃) $\delta -5.4 (\times 2)$, 13.6, 15.5, 16.6, 19.2, 20.2, 25.0, 25.9 (\times 3), 27.3, 28.1, 31.5, 32.3, 33.50, 35.7, 36.0, 36.1, 36.4, 38.0, 47.9, 50.5, 54.5, 56.4, 60.4, 68.4, 69.8, 71.6, 78.2, 79.0, 116.2, 118.4 (q), 126.9 (\times 2), 127.1, 127.3, 127.5 (\times 2), 128.1 (\times 2), 128.3 (\times 2), 139.0, 139.5, 153.1; HR EIMS *m*/*z* 860.4648 (calcd 860.4693 for C₄₈H₇₁F₃O₆SSi).

(25*R*)-26-[(*tert*-Butyldimethylsilyl)oxy]-3β,6β-bis(benzyloxy)-5α-cholest-15-ene (13). To a solution of 12 (0.038 g, 0.045 mmol) in THF (2 mL) were added LiCl (0.008 g, 0.200 mmol), Pd(PPh₃)₄ (0.002 g, 0.002 mmol), and Bu₃SnH (0.018 mL, 0.062 mmol). The reaction mixture was refluxed for 16 h, then quenched with water (1 mL), concentrated in vacuo to remove the excess of THF, and extracted with petroleum ether. The organic layer was washed with a solution of NH₄OH (10% in water), and then with brine and finally dried (Na₂SO₄) and concentrated in vacuo. The residue was flash chromatographed (silica gel, 0–1% diethyl ether in petroleum ether) to give 13 as a colorless oil (0.016 g, 50%). 13: [α]_D –6.5 (*c* = 1.8, CHCl₃); ¹H NMR (250 MHz, CDCl₃)

13: $[\alpha]_{\rm D} - 6.5$ (c = 1.8, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.07 (6 H, s, (CH₃)₂-Si), 0.77 (3 H, s, Me-18), 0.88 (3 H, d, J = 6.7 Hz, Me-27), 0.91 (9 H, s, (CH₃)₃-C), 0.92 (3 H, d, J = 6.0 Hz, Me-21), 1.10 (3 H, s, Me-19), 3.36 (1 H, m, H-3), 3.37 (1 H, dd, J = 9.7, 6.5 Hz, H-26), 3.47 (1 H, J = 9.7, 5.9 Hz, H'-26), 3.49 (1 H, bs, H-6), 4.35 (1 H, d, J = 12.2 Hz, CHPh), 4.57 (2 H, s, CH₂Ph), 4.65 (1 H, d, J = 12.2 Hz, CHPh), 5.79 (1 H, bd, J = 6.2 Hz, H-15 or H-16), 5.83 (1 H, bd, J = 5.8 Hz, H-15 or H-16), 7.35 (10 H, m, C₆H₃); ¹³C NMR (62.5 MHz, CDCl₃) δ -5.3 (× 2), 12.8, 15.6, 16.7, 18.3, 18.5, 20.8, 23.8, 26.0 (× 3), 28.0, 28.3, 32.2, 32.5, 33.5, 35.2, 35.7, 36.1, 36.5, 37.7, 38.3, 48.1, 49.6, 55.1, 61.5, 62.4, 68.5, 69.8, 71.3, 78.5, 79.3, 126.9 (× 2), 127.1, 127.3, 127.5 (× 2), 128.1 (× 2), 128.3 (× 2), 130.8, 133.8, 139.2, 139.7; HR EIMS *m*/*z* 712.5238 (calcd 712.5251 for C₄₇H₇₂O₃Si).

(25*R*)-3 β ,6 β -Bis(benzyloxy)-5 α -cholest-15-en-26-ol (14). To a solution of 13 (0.037 g, 0.052 mmol) in THF (0.5 mL) was added Bu₄NF (0.1 mL, 1 M in THF, 0.1 mmol). After 16 h the reaction mixture was quenched with water (1 mL), concentrated in vacuo to remove the excess of THF, and extracted with ethyl acetate. The organic layer was dried (Na₂SO₄) and concentrated in vacuo, and the residue was flash chromato-

graphed (silica gel, 5-20% ethyl acetate in petroleum ether) to give **14** as a colorless oil (0.017 g, 54\%).

14: $[\alpha]_{\rm D} - 27.1 \ (c = 1.0, \text{CHCl}_3)$; ¹H NMR (400 MHz, CDCl}3) δ 0.75 (3 H, s, Me-18), 0.90 (3 H, d, J = 6.7 Hz, Me-27), 0.92 (3 H, d, J = 6.0 Hz, Me-21), 1.08 (3 H, s, Me-19), 2.20 (1 H, bd, J = 13.7 Hz, H-7 β), 3.39 (1 H, m, H-3), 3.42 (1 H, dd, J =9.7, 6.7 Hz, H-26), 3.47 (1 H, bs, H-6), 3.49 (1 H, J = 9.7, 5.9Hz, H'-26), 4.35 (1 H, d, J = 12.2 Hz, CHPh), 4.56 (2 H, s, CH₂Ph), 4.64 (1 H, d, J = 12.2 Hz, CHPh), 5.74 (1 H, bd, J =6.2 Hz, H-15 or H-16), 5.81 (1 H, bd, J = 5.8 Hz, H-15 or H-16), 7.29 (10 H, m, C₆H₃); ¹³C NMR δ (100 MHz, CDCl₃) 12.8, 15.6, 16.5, 18.4, 20.7, 23.8, 28.0, 28.2, 32.1, 32.4, 33.5, 35.1, 35.8, 36.1, 36.4, 37.7, 38.2, 48.0, 49.6, 55.0, 61.5, 62.3, 68.5, 69.7, 71.2, 78.4, 79.2, 126.8 (× 2), 127.0, 127.3, 127.6 (× 2), 128.1 (× 2), 128.3 (× 2), 130.9, 133.7, 139.1, 139.7; HR EIMS *m*/*z* 598.4377 (calcd 598.4386 for C₄₁H₅₈O₃).

(25*R*)-3 β ,6 β -Bis(benzyloxy)-15 β ,16 β -epoxy-5 α -cholestan-26-ol (15). To a solution of 14 (0.007 g, 0.012 mmol) in methylene chloride (0.5 mL) at 0 °C was added MCPBA (0.004 g, 0.024 mmol). After 5 h the reaction mixture was quenched with a saturated solution of Na₂SO₃ (1 mL), extracted with methylene chloride, dried (Na₂SO₄), and concentrated in vacuo. The residue was flash chromatographed (silica gel, 0–2% methanol in methylene chloride) to give 15 as a colorless oil (0.004 g, 57%).

15: $[\alpha]_{\rm D}$ -19.7 (*c* = 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.92 (3 H, s, Me-18), 0.93 (6 H, d, *J* = 7.0 Hz, Me-27 and Me-21), 1.05 (1 H, d, *J* = 11.6 Hz, H-14), 1.08 (3 H, s, Me-19), 2.26 (1 H, bd, *J* = 13.7 Hz, H-7 β), 3.06 (1 H, bs, H-15 or H-16), 3.20 (1 H, d, *J* = 3.2 Hz, H-15 or H-16), 3.36 (1 H, m, H-3), 3.44 (1 H, dd, *J* = 10.4, 6.7 Hz, H-26), 3.50 (1 H, bs, H-6), 3.51 (1 H, *J* = 10.4, 6.0 Hz, H'-26), 4.43 (1 H, d, *J* = 12.2 Hz, *CH*Ph), 4.55 (2 H, s, *CH*₂Ph), 4.59 (1 H, d, *J* = 12.2 Hz, *CH*Ph), 7.29 (10 H, m, C₆*H*₅); ¹³C NMR δ (100 MHz, CDCl₃) 15.2, 16.4, 18.5, 20.3, 23.9, 27.2, 28.2, 29.6, 32.3 (× 2), 33.3, 35.3, 35.7, 36.1, 36.9, 37.2, 38.1, 48.0, 53.7, 54.7, 58.8, 59.3 (× 2), 64.1, 68.4, 69.8, 71.6, 78.3, 79.4, 126.9 (× 2), 127.0, 127.3, 127.5 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.8; HR EIMS *m*/*z* 614.4306 (calcd 614.4335 for C₄₁H₅₈O₄).

(25*R*)-26-[(*tert*-Butyldimethylsilyl)oxy]-3 β ,6 β -bis(benzyloxy)-16-[(trimethylsilyl)oxy]-5 α -cholest-15-ene (16). To a solution of 11 (0.200 g, 0.275 mmol) in THF (2 mL) at -78°C was added LiN(SiMe₃)₂ (0.55 mL, 1.0 M in THF, 0.55 mmol). After 1 h Me₃SiCl (0.10 mL, 0.83 mmol) was added, and after additional 0.3 h, the mixture was allowed to warm to room temperature. The reaction was then quenched by addition of water, concentrated in vacuo to remove the excess of THF, and extracted with petroleum ether. The organic phase was dried (Na₂SO₄) and concentrated in vacuo to give crude 16 as a colorless oil (0.240 g). 16 was used in the next step without further purification.

16: ¹H NMR (250 MHz, CDCl₃) δ 0.07 (6 H, s, (CH₃)₂-Si), 0.24 (9 H, s, (CH₃)₃-Si), 0.91 (3 H, s, Me-18), 0.92 (3 H, d, J = 6.7 Hz, Me-27), 0.95 (9 H, s, (CH₃)₃-C), 1.09 (3 H, d, J = 6.0 Hz, Me-21), 1.18 (3 H, s, Me-19), 3.37 (1 H, m, H-3), 3.38 (1 H, dd, J = 9.8, 6.7 Hz, H-26), 3.48 (1 H, dd, 9.8, 5.9 Hz, H'-26), 3.49 (1 H, bs, H-6), 4.39 (1 H, d, J = 12.2 Hz, CHPh), 4.58 (1 H, s, CH₂Ph), 4.61 (1 H, bs, H-15), 4.66 (1 H, d, J = 12.2 Hz, CHPh), 7.33 (10 H, m, C₆H₃); ¹³C NMR (62.5 MHz, CDCl₃) δ -5.3 (× 2), -5.4 (× 3), 13.6, 15.6, 16.7, 18.3, 19.4, 20.4, 22.6, 25.2, 26.0 (× 3), 28.1, 28.3, 32.1, 32.4, 33.7, 35.3, 35.9, 36.1, 36.7, 36.9 38.1, 48.1, 49.1, 54.8, 56.6, 61.3, 68.6, 69.8, 71.4, 78.5, 79.5, 101.2, 126.9 (× 2), 127.0, 127.3, 127.5 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.8, 158.7.

(25*R*)-3 β ,6 β -Bis(benzyloxy)-5 α -cholestane-15 α ,26-diol-16-one (18). To a solution of 16 (0.220 g 0.275 mmol) in CH₂-Cl₂ at 0 °C was added dimethyldioxirane (3.3 mL 0.1 M in acetone, 0.33 mmol). After 1 h the mixture was concentrated in vacuo, and the residue was dissolved in acetone (4 mL). Camphorsulfonic acid (0.01 g, 0.04 mmol) was added, and the mixture was left at 4 °C for 16 h. The reaction mixture was then diluted with water (2 mL), concentrated in vacuo to remove the excess of acetone, extracted with chloroform, dried (Na₂SO₄), and concentrated in vacuo. The crude residue was purified by flash chromatography (0-1% methanol in chloroform) to give **18** (0.123 g, 71\%, overall yield) as a white solid.

18: mp 53–55 °C; $[\alpha]_D$ –31.2 (c = 1.0, CHCl₃); ¹H NMR (250 MHz, CDCl₃) δ 0.88 (3 H, s, Me-18), 0.92 (3 H, d, J = 6.7 Hz, Me-27), 1.01 (3 H, d, J = 6.0 Hz, Me-21), 1.12 (3 H, s, Me-19), 3.37 (1 H, m, H-3), 3.39 (1 H, dd, J = 9.8, 6.7 Hz, H-26), 3.49 (1 H, dd, J = 9.8, 5.9 Hz, H'-26), 3.50 (1 H, bs, H-6), 3.59 (1 H, d, J = 11.5 Hz, H-15), 4.39 (1 H, d, J = 12.2 Hz, CHPh), 4.57 (2 H, bs, CH₂Ph), 4.63 (1 H, d, J = 12.2 Hz, CHPh), 7.33 (10 H, m, C₆H₅); ¹³C NMR (62.5 MHz) δ 15.7, 15.8, 16.5, 19.3, 20.3, 24.4, 28.1, 29.8, 31.2, 32.2, 33.1, 34.9, 35.1, 35.5, 36.0, 38.1, 39.1, 39.7, 47.4, 53.9, 56.1, 64.5, 68.1, 69.7, 71.3, 76.6, 77.2, 78.2, 79.1, 126.8 (× 2), 127.2, 127.4 (× 2), 128.0 (× 2), 128.2 (× 2), 139.0, 139.8, 213.1; HR EIMS *m*/*z* 630.4256 (calcd 630.4284 for C₄₁H₅₈O₅).

(25*R*)-3 β ,6 β -Bis(benzyloxy)-5 α -cholestane-15 α ,16 α ,26triol (19) and (25*R*)-3 β ,6 β -Bis(benzyloxy)-5 α -cholestane-15 α ,16 β ,26-triol (20). (a) Reduction with LiAlH₄. To a solution of 18 (0.042 g, 0.67 mmol) in diethyl ether (2 mL) at 0 °C was added LiAlH₄ (0.2 mL, 1.0 M in THF, 0.2 mmol). The reaction mixture was stirred for 0.2 h and quenched with ethyl acetate (0.5 mL) and NH₄OH (0.2 mL, 30% aqueous solution). Filtration through a short pad of Celite and concentration in vacuo gave a crude residue which was purified by flash chromatography (0–1% methanol in chloroform) to give 19 as colorless oil (0.017 g, 40%) and 20 (0.021 g, 50%) as white solid.

(b) Reduction with BH₃·SMe₂. To a solution of **18** (0.022 g, 0.35 mmol) in THF (3 mL) at 0 °C was added BH₃·SMe₂ (0.055 mL, 2 M in THF, 0.11 mmol). After 2 h the mixture was quenched with 0.1 M HCl, concentrated in vacuo to remove the excess of THF, extracted with chloroform (6 mL), and washed with a saturated solution of NaCl. The organic phase was then dried (Na₂SO₄) and concentrated to give a crude residue which was purified by flash chromatography (0–1% methanol in chloroform) to give **20** (0.013 g, 58%).

19: $[\alpha]_D$ -6.3 (c = 1.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.70 (3 H, s, Me-18), 0.91 (3 H, d, J = 7.0 Hz, Me-27), 0.95 (3 H, d, J = 6.6 Hz, Me-21), 1.07 (3 H, s, Me-19), 2.43 (1 H, bd, J = 14.3 Hz, H-7 β), 3.36 (1 H, m, H-3), 3.47 (3 H, m, H-6 and H₂-26), 3.64 (1 H, dd, J = 8.2, 8.0 Hz, H-15), 3.80 (1 H, dd, J = 7.2, 7.0 Hz, H-16), 4.37 (1 H, d, J = 12.2 Hz, CHPh), 4.56 (2 H, m, CH₂Ph), 4.60 (1 H, d, J = 12.2 Hz, CHPh), 7.33 (10 H, m, C₆H₃); ¹³C NMR (100 MHz, CDCl₃) δ 15.3, 15.8, 16.6, 19.2, 20.5, 23.8, 28.3, 30.2, 32.4, 33.5, 33.9, 35.2, 35.6, 35.7, 36.0, 38.5, 39.8, 40.3, 47.5, 54.2, 59.8, 63.6, 68.4, 69.8, 71.4, 72.8, 74.8, 78.4, 79.4, 126.9 (× 3), 127.4, 127.6 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.9; HR FABMS *m/z* 633.4492 (calcd 633.4519 for C₄₁H₆₁O₅, [M + H]⁺).

20: mp 61–63 °C; $[\alpha]_D$ –1.5 (c = 2.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.88 (3 H, s, Me-18), 0.89 (3 H, d, J = 7.0 Hz, Me-27), 0.95 (3 H, d, J = 6.6 Hz, Me-21), 1.07 (3 H, s, Me-19), 2.29 (1 H, bd, J = 14.3 Hz, H-7 β), 3.36 (1 H, m, H-3), 3.45 (3 H, m, H-6 and H₂-26), 3.75 (1 H, dd, J = 10.0, 1.6 Hz, H-15),

4.06 (1 H, dd, J = 7.4, 1.6 Hz, H-16), 4.37 (1 H, d, J = 12.2 Hz, *CH*Ph), 4.55 (2 H, bs, *CH*₂Ph), 4.60 (1 H, d, J = 12.2 Hz, *CH*Ph), 7.33 (10 H, m, C₆H₃); ¹³C NMR (100 MHz, CDCl₃) δ 14.6, 15.8, 16.7, 17.9, 20.6, 23.5, 28.2, 29.3, 30.2, 32.3, 33.2, 35.1, 35.4, 35.8, 35.9, 38.4, 40.2, 43.8, 47.5, 54.2, 58.6, 59.9, 68.3, 69.7, 71.4, 78.3, 79.3, 82.4, 83.9, 126.9 (× 2), 127.0, 127.3, 127.5 (× 2), 128.1 (× 2), 128.3 (× 2), 139.1, 139.9; HR FABMS *m*/*z* 633.4487 (calcd 633.4519 for C₄₁H₆₁O₅, [M + H]⁺).

(25*R*)-5 α -Cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol (1a). To a solution of 20 (0.068 g, 0.107 mmol) in absolute ethanol (2.5 mL) was added palladium on activated carbon (7 mg). The flask was evacuated (20 Torr) and flushed with hydrogen three times. The reaction mixture was then stirred vigorously under an atmosphere of hydrogen for 5 h. The reaction mixture was filtered through a pad of silica gel and concentrated to give 1a (0.048 g, 0.104 mmol, 100%).

1a: mp 133–135 °C; $[\alpha]_D$ +29.5 (*c* = 2.3, MeOH); ¹H NMR (400 MHz, CD₃OD₃) δ 0.93 (3 H, d, J = 6.9 Hz, Me-27), 0.95 (3 H, s, Me-18), 0.99 (3 H, d, J = 6.6 Hz, Me-21), 1.07 (3 H, s, Me-19), 2.18 (1 H, bd, J = 14.3 Hz, H-7 β), 3.36 (1 H, dd, J =10.5, 6.8 Hz H-26), 3.44 (1 H, J = 10.5, 5.9 Hz H'-26), 3.57 (1 H, m, H-3), 3.77 (1 H, bs, H-6), 3.78 (1 H, dd, overlapped with H-6, H-15), 4.00 (1 H, dd, J = 7.6, 2.0 Hz, H-16), 4.37 (1 H, d, J = 12.2 Hz, CHPh), 4.55 (1 H, bs, CH₂Ph), 4.60 (1 H, d, J = 12.2 Hz, CHPh), 7.33 (10 H, m, C₆H₅); ¹³C NMR (100 MHz, CD₃OD₃) & 15.0 (C-18), 16.3 (C-19), 17.1 (C-27), 18.6 (C-21), 21.9 (C-11), 24.8 (C-23), 30.9 (C-20), 31.2 (C-8), 32.2 (C-2), 34.7 (C-24), 36.3 (C-4), 36.6 (C-10), 36.9 (C-25), 37.3 (C-22), 39.8 (C-1), 40.6 (C-7), 41.8 (C-12), 44.7 (C-13), 48.8 (C-5), 55.7 (C-9), 60.0 (C-17), 61.1 (C-14), 68.6 (C-26), 72.4 (C-3), 72.5 (C-6), 82.9 (C-16), 85.0 (C-15); HR FIBMS m/z 453.3577 (calcd 453.3580 for $C_{27}H_{49}O_5$, $[M + 1]^+$).

Acknowledgment. This work has been supported by the European Commission, MAST-III program (contract no. MAS3-CT95-0032), by MURST (40%), and by CNR (Rome). The mass spectra were obtained from the "Servizio di Spettrometria di Massa del CNR e dell'Università di Napoli" and from the CRIAS (Centro Interdipartimentale di Analisi Strumentale) dell'Università di Napoli "Federico II"; the staffs are gratefully acknowledged.

Supporting Information Available: ¹H NMR spectra of compounds **1a** and **3–20**, ¹³C NMR spectra of compounds **1a**, **3–11**, **13–15**, and **18–20**, and two-dimensional spectra of compounds **15**, **18**, and **19** (42 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

JO980266C